# THE STUDY OF NATURAL POLYSACCHARIDES: ORGANOLEPTIC, PHYSICAL, CHEMICAL, MICROBIOLOGICAL PROPERTIES, AND THERMODYNAMIC CHARACTERISTICS OF AQUEOUS SOLUTIONS

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Abstract: Natural polysaccharides is a promising raw material for biodegradable polymers production. To properly select sources of their origin would require conformity with safety standards in force for the similar products. This work conducts a study with a view to examine the extent to which the organoleptic, physical and chemical properties of the polysaccharides in question, as well as their safety indicators conform with the provisions of GOST 16280-2002, CAS 11114-20-8 and CAS 9004-65-3. It is determined that the samples selected, namely, agar-agar produced by Panreac (Germany) and Helicon (USA), carrageenan produced by Boc Sciences (USA) and Newgreen Pharmchem Co. (China) and hydroxypropylmethyl cellulose produced by Acros (Belgium) and Ashland Aqualon Functional Ingredients (USA), meet safety requirements for this type of raw material. Simultaneously, we employ high-sensitivity differential scanning calorimetry method to analyze thermodynamic characteristics of natural polysaccharides' aqueous solutions. It is shown that gels based on agar-agar have high melting temperature and can withstand temperature rise; after, their microscopic structure is not affected by thermal history, and the gel system remains stable. Thermodynamically, aqueous solution of kappa-carrageenan forms more stable gel than iota-carrageenan; strength and melting point of kappa-carrageenan gels can be adjusted by changing the concentration of potassium chloride in the solution. Aqueous solutions of hydroxypropylmethylcellulose are thermodynamically stable. The experimental data obtained confirm the feasibility of natural polysaccharides application in the production of biodegradable polymers.

Keywords: natural polysaccharides, agar-agar, carrageenan, hydroxypropyl methylcellulose, biodegradable polymers

#### INTRODUCTION

Nowadays, it is hard to find the sphere of human activity that does not employ polymer materials. As polymers production from petrochemicals increases due to their high demand, chemical stability of the synthetic polymers remains the problem at hand. The polymers can withstand well physical and chemical natural factors such as light, heat, humidity, atmospheric oxygen, as well as microbiological natural factors without any obvious signs of decomposition for decades. Polymers and their traces stay in the environment for a long time causing a substantial environmental damage [1, 2].

Similar problems occur upon synthetic polymers' disposal. Most synthetic materials emit poison gas during incineration, and contaminate groundwater in landfill as they do not rot down. Some countries (Taiwan, Germany, Ireland, and South Africa) now banned or limited the plastic bags usage. In addition, the declining oil and gas reserves tend to increase the cost of the materials [3].

Hence, it becomes important to produce modern polymeric materials from naturally occurring polymers,

e.g. starch, chitosan, agar-agar, cellulose, carrageenan, pectin, etc. that will enter the biogeochemical cycle, and are environmentally safe due to their structure.

Unlike other plastics, biodegradable polymers will decompose into the low-molecular substances to take part in metabolism of protozoan species by acting chemically, physically, or biologically. Admittedly, this property of new materials will resolve the issue of waste disposal [4, 5].

Biodegradable polymers are produced by polymerizing the raw materials of biological origin. The latter are either extracted from plants and animals or synthesized using the commercial raw-materials processing technologies [6].

Biodegradable polymers offer substantial benefits in biocompatibility and environmental safety. Owing to that fact, they should see increasing application in medicine (disposable surgical products), pharmaceutical industry (medicines manufacturing), food industry (disposable packaging and tableware), and in agriculture (degradable films and bags) [7].

In simplistic terms, natural polysaccharidescontaining products belong to gel-like systems that

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have complex distribution of water molecules. Gels are multi-component systems consisting of high-molecular substance (-s) and low-molecular liquid (water) [8].

Functional properties of polysaccharides suspensions depend on the pH value, temperature, particles sizes, surface structure, as well as composition such as salt content. Consequently, to practically introduce polysaccharides in actual multi-component systems would involve the assessment of the wide range of physical-chemical and functional properties [9, 10].

Therefore, it is desirable to comprehensively describe natural polysaccharides that are used in production of biodegradable polymers, their organoleptic, physical-chemical, thermodynamic properties, as well as safety.

# **OBJECTS AND METHODS OF STUDY**

The objects of study includ the following polysaccharides: agar-agar, carrageenan, hydroxypropyl methylcellulose (HPMC). The natural polysaccharides under consideration display a range of suitable properties; as raw materials they have a potential to improve biodegradable polymers production processes.

Agar-agar is cold water-insoluble. It fully dissolves only at temperatures between 95 and 100°C. The heated solution is transparent, and of limited viscosity. When cooled down to 35-40°C agar-agar forms clear and strong thermodynamically reversible gel. When heated to 85-95°C it melts to become a liquid solution and then, transforms back to gel at 35-40°C.

Carrageenans are large, highly flexible molecules that curl forming helical structures. This permits the formation of a variety of gels at room temperature. The primary differences that affect properties essential to each carrageenans' class are the number and position of the ester sulfate groups on the repeating galactose units. Higher levels of ester sulfate lower the solubility temperature of the carrageenan producing the lower strength gels, and thus, contribute to gel inhibition (lambda carrageenan).

Among other implications, hydroproxypropyl methylcellulose is a stabilizer, an emulsifier and a thickening agent added to food to preserve and increase its viscosity.

For our study, we selected the following natural polysaccharides:

- agar-agar (Panreac, Germany);

- agar-agar (Helicon, USA);
- kappa-carrageenan (Boc Sciences, USA);
- iota-carrageenan (Newgreen Pharmchem Co., China);

- hydroproxypropyl methylcellulose (Acros, Belgium);

- hydroproxypropyl methylcellulose (Ashland Aqualon Functional Ingredients, USA).

To analyze the organoleptic, physical-chemical and microbiological properties tested, we referred to the following regulatory documents applicable to the natural polysaccharides under investigation:

- GOST 16280-2002 Food grade agar. Specifications;

- carrageenans. CAS 11114-20-8;

- hydroxypropyl methylcellulose. CAS 9004-65-3.

We employed the DSC 204 F1 high sensitivity differential scanning calorimeter (Phoenix®, Germany) to determine thermodynamic quantities of polysaccharides aqueous solutions.

# **RESULTS AND DISCUSSION**

To study the above-listed raw materials for purposes of practical application in biodegradable polymers production, we examined organoleptic, physical-chemical indicators, analyzed chemical and microbiological safety, and thermodynamic quantities of their aqueous solutions

Table 1 shows the results of examining agar-agar organoleptic properties from the producers selected.

Agar-agar organoleptic analysis indicates that the samples tested are in the form of homogenous lightcream powder with gray hue, without off-odors and off-taste; visible foreign impurities to the unaided eye not found.

Table 2 illustrates data obtained when testing physical-chemical properties of the agar-agar samples investigated.

The above-listed data proves that physical-chemical properties of both agar-agar samples satisfy the requirements.

Table 3 shows outcome of testing the microbiological and chemical safety of agar-agar, heavy metals content and microbiological indicators included.

Table 3 proves chemical and microbiological safety indicators of both agar-agar samples to satisfy the agaragar GOST 16280-2002 current requirements. The samples do not contain coliform bacteria, proteobacteria, and pathogenic microorganisms.

Based on the analysis of organoleptic and physicalchemical properties, as well as chemical and microbiological safety indicators it can be concluded that each of the agar-agar samples selected satisfies GOST 16280-2002 overall requirements and can be recommended for biodegradable polymers production.

Then, we assessed the organoleptic indicators of the carrageenans selected (USA, China). The experimental results are shown in Table 4. Analyzing the data we referred to the international food standards CODEX Alimentarius provisions, a CAS 11114-20-8 standard for carrageenans.

The results revealed that these caraggenan samples form homogenous fine powder of yellowish-white color without foreign taste or odor.

Table 5 presents data obtained upon examination of carrageenan physical-chemical properties.

As per data shown in Table 5, the carrageenan samples concerned display satisfactory results in regards to mass fraction of moisture. Within the limits of the norm are mass fractions of ash and sulfates. The samples are ethanol-insoluble, water-soluble at a temperature of 80°C therewith forming viscous transparent or slightly opalescent solutions.

Table 6 shows an outcome of examining the carrageenan chemical and microbiological safety, heavy metals content and microbiological indicators.

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dry matter 0.85%

dry matter 0.85%

Impurities

Taste, gel w/mass fraction of

Table 6 proves that chemical and microbiological safety indicators of both carrageenan samples meet 11114-20-8 requirements of the current CAS international food standards. The samples do not exhibit coliform bacteria and pathogenic microorganisms.

Similarly, in Table 7 we considered organoleptic properties of hydroproxypropyl methylcellulose (HPMC) from the producers selected (Belgium, USA) compare the results against international to standards CODEX Alimentarius, food а CAS 9004-65-3 standard for HPMC.

No off-taste

Shall not be permitted

No off-taste

Not found

	Sampl		
Indicator	The Panreac agar-agar (Germany)	The Helicon agar-agar (USA)	GOST 16280-2002 provisions
Appearance	Homogenous powder	Homogenous powder	Grist, granules, flakes, powder, lamella, film
Color	Light-cream with gray hue	Cream with gray hue	Light-cream to dark-cream, gray hue permitted
Odour, gel w/mass fraction of	No off-odors, weak odor of	No off-odors	No off-odors

Table 1. The results of agar-agar organoleptic indicators' analysis

Table 2. T	he results	obtained	through	analysis	of agar-agar	physical	l-chemical	indicators
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algae

No off-taste

Not found

	San	nple	GOST 16280-2002
Indicator	The Panreac agar-agar (Germany)	The Helicon agar-agar (USA)	provisions
Color, gel w/mass fraction of dry agar 0.85%, light transmission %	$62.0 \pm 3.1$	$65.0 \pm 3.3$	Over 60.0
Strength, gel w/mass fraction of dry agar 0.85% and sugar 70%, g	$1650\pm165$	$1600 \pm 160$	Over 1600
Lowered strength after heating for 2 hrs, gel w/mass fraction of dry agar 0.85%, %	$10.0 \pm 1.0$	$9.0 \pm 0.9$	Under 10.0
Melting temperature, gel w/mass fraction of dry agar 0.85%, °C	$82.0 \pm 8.2$	85.0 ± 8.5	Over 80.0
Gel-forming temperature, agar aqueous solution gel w/mass fraction of dry agar 0.85%, °C	32.0 ± 3.2	32.0 ± 3.2	Over 30.0
Gel-forming temperature, agar aqueous solution w/mass fraction of dry agar 0.85% and sugar 70%, °C	$40.0 \pm 4.0$	$42.0 \pm 4.2$	Under 42.0
Mass fraction of water, %	$16.0\pm1.6$	$17.5 \pm 1.8$	Under 18.0
Mass fraction of ash, %	$4.0 \pm 0.4$	$4.2 \pm 0.4$	Under 4.5
Iodine presence	Not found	Not found	Shall not be permitted
Mass fraction of hot-water insoluble substances, %	$0.20 \pm 0.02$	$0.30 \pm 0.03$	Under 0.40

Table 3	The results of the agar	-agar chemical and	l microbiological	safety analysis
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	Sam	Sample		
Indicator	The Panreac agar-agar (Germany)	The Helicon agar-agar (USA)	GOST 16280-2002 provisions	
Lead, mg/kg	$0.15 \pm 0.01$	$0.08 \pm 0.01$	Under 2.00	
Cadmium, mg/kg	Under detection limit	$0.010 \pm 0.005$	Under 0.030	
Arsenic, mg/kg	$0.010 \pm 0.005$	$0.010 \pm 0.005$	Under 0.500	
Mercury, mg/kg	Under detection limit	Under detection limit	Under 0.02	
QMAFAnM, CFU/g	$0.9 \cdot 10^2$	$3.2 \cdot 10^2$	Under $1.0 \cdot 10^3$	
Coliform bacteria, CFU/g	Not found	Not found	Shall not be permitted	
Proteobacteria	Not found	Not found	Shall not be permitted	
Pathogenic microorganisms	Not found	Not found	Shall not be permitted	

# **Table 4.** The results of analyzing the carrageenans' organoleptic indicators

		CAS 11114-20-8	
Indicator	The Boc Sciences kappa- carrageenan (USA)	The Newgreen Pharmchem Co. iota- carrageenan (China)	provisions
Appearance and consistency	Fine powder	Fine powder	Fine powder
Odor	No	No	No
Color	Yellowish-white	Yellowish-white	From yellowish-white to brown
Taste	No	No	No

# Table 5. Physical-chemical quantities observed when testing carrageenans

	Sar	nple	CAS 11114-20-8	
Indicator	The Boc Sciences kappa- carrageenan (USA)	The Newgreen Pharmchem Co. Iota-carrageenan (China)	provisions	
Mass fraction of moisture, %	$10.5 \pm 1.0$	$11.0 \pm 1.1$	Under 12.0	
Suspension pH 1 : 100	9.5 ± 1.0	$10.0 \pm 1.0$	8.0-11.0	
1.5% solution viscosity at 75°C, cP	$6.5 \pm 0.3$	5.5 ± 0.3	Over 5.0	
Mass fraction of sulfates, %	25.0 ± 2.5	18.0 ± 1.8	15.0-40.0	
Mass fraction of ash, %	$15.0 \pm 1.5$	17.5 ± 1.8	15.0-40.0	
Mass fraction of ash, acid-insoluble, %	$0.80 \pm 0.08$	$0.50 \pm 0.05$	Under 1.00	
Mass fraction of the substance, acid- insoluble, %	$1.0 \pm 0.1$	1.5 ± 0.2	Under 2.0	
Mass fraction of solvent (ethanol, isopropanol or methanol), %	Not found	$0.050 \pm 0.005$	Under 0.100	

# Table 6. The results of carrageenan chemical and microbiological safety analysis

	Sam	Sample		
Indicator	The Boc Sciences kappa- carrageenan (USA)	The Newgreen Pharmchem Co. carraggeenan (China)	CAS 11114-20-8 provisions	
Lead, mg/kg	$0.50 \pm 0.03$	$1.10 \pm 0.06$	Under 5.0	
Arsenic, mg/kg	$0.20 \pm 0.01$	$0.50 \pm 0.03$	Under 3.0	
Cadmium, mg/kg	$0.10 \pm 0.01$	$0.10 \pm 0.01$	Under 2.0	
Mercury, mg/kg	Not found	Not found	Under 1.0	
QMAFAnM, CFU/g	$1.0 \cdot 10^2$	$1.5 \cdot 10^{1}$	Under $5.0 \cdot 10^3$	
Pathogenic microorganisms, salmonella incl.	Not found	Not found	Shall not be permitted	
Coliform bacteria, CFU/g	Not found	Not found	Shall not be permitted	

# Table 7. The results obtained when analyzing the organoleptic indicators of hydroproxypropyl methylcellulose

	S	CAS 9004-65-3	
Indicator	The Acros HPMC (Belgium)	The Ashland Aqualon Functional Ingredients HPMC (USA)	provisions
Appearance and consistency	Hygroscopic powder	Hygroscopic powder	Hygroscopic powder, granules or fibres
Odor	No	No	No
Color	White	White	From white to off white
Taste	No	No	No

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Hydroproxypropyl methylcellulose organoleptic analysis revealed that the samples investigated are in the form of hygroscopic powder of white color without foreign taste or odor.

The data obtained through testing the physicalchemical indicators of HPMC samples are shown in Table 8.

Experimentally, the physical-chemical indicators of hydroproxypropyl methylcellulose samples in question display acceptable results in regards to mass fraction of moisture, sulfated ash and pH solution value contents.

Table 9 considers the results of hydroproxypropyl methylcellulose chemical and microbiological safety analysis, as well as heavy metals content and microbiological indicators.

Table 9 proves the conformance of both HPMC samples with the current international food standard CAS 9004-65-3 requirements in terms of chemical and microbiological safety. The HPMC samples under investigation do not contain coliform bacteria and pathogenic microorganisms.

Further, we examined thermodynamic characteristics of the foregoing polysaccharides aqueous solutions.

One of the main methods to do so for biopolymers aqueous solutions and dispersions is high sensitivity differential scanning calorimetry (DSC). The technique is used to measure temperatures and enthalpies of conformational change in biopolymer solutions and dispersions, and to qualitatively describe gel-forming capacity of naturally-occurring polymers.

It is well-known that agar-agar is sparingly soluble in cold water. When swelling, agar-agar forms colloidal solution; the latter produces a strong gel. An initial agar-agar gel-forming concentration of 0.5% launches intermolecular bonding in the biopolymer solution. Agar-agar creates gel by forming and aggregating double helix, regardless of the proportions of cation and low-molecular weight compounds in the solution.

Fig. 1 shows DSC thermogram for agar-agar aqueous dispersions. Thermodynamic characteristics of the same are displayed in Table 10.

After analyzing the Fig. 1 and Table 10, we were able to see a broad endothermic phase transition from 60 to 95°C in agar-agar 0.5% aqueous solutions, which is inherent to changes from double helix to random coil conformation. An endothermic transition value Tt of 81.3°C was found for the German sample and 84.5°C for the American sample.

Therefore, agar-agar-based gels are characterized by high melting temperature (over 80°C) and by ability to withstand the temperature rise. When re-scanned, agar-agar aqueous dispersions proved conformational change to be reversible. Heating transforms agar-agar gels to liquid, and after cooling solidifies them back. The microscopic structure of these gels is not affected (within certain range) by thermal history of the sample. This property is important when creating a stable gel system under the variable environmental conditions.

Table 8. The results of HPMC physical-chemical indicators' analysis

		CAS 9004-65-3	
Indicator	The Acros HPMC (Belgium)	The Ashland Aqualon Functional Ingredients HPMC (USA)	provisions
Mass fraction of moisture, %	$8.0 \pm 0.8$	$8.5 \pm 0.9$	Under 10.0
Solution pH 1 : 100	6.5	7.0	5.0-8.0
Mass fraction of sulfated ash, %	1.2 ± 0.1	$1.5 \pm 0.1$	Under 1.5 for samples with a viscosity of 50 cP and over. Under 3.0 for samplea with a viscosity of 50 cP and under

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Table 9. The results of hydroproxypropyl	i meliivicenuiose chemicai	i and iniciopiopogical	salely mulcalors analysis

	Sa	CAS 9004-65-3	
Indicators	The Acros (Belgium)	The Ashland Aqualon Functional Ingredients HPMC (USA)	provisions
Lead, mg/kg	$0.10 \pm 0.01$	$0.25 \pm 0.01$	Under 2.0
Arsenic, mg/kg	$0.30 \pm 0.01$	$0.50 \pm 0.03$	Under 3.0
Cadmium, mg/kg	$0.25 \pm 0.01$	$0.18 \pm 0.01$	Under 2.0
Mercury, mg/kg	Not found	Not found	Under 1.0
Propylene chlorohydrin, mg/kg	Not found	Not found	Under 1.0
QMAFAnM, CFU/g	Not found	Not found	Under $5.0 \cdot 10^3$
Pathogenic microorganisms, salmonella incl.	Not found	Not found	Shall not be permitted
Coliform bacteria, CFU/g	Not found	Not found	Shall not be permitted

### Table 10. Thermodynamic characteristics of melting the agar-agars' aqueous dispersions

Sample	T <sub>t</sub> , ℃	$\Delta H_t$ , J/g
The Panreac agar-agar	$81.3 \pm 4.1$	$24.5 \pm 1.2$
The Helicon	$84.5 \pm 4.2$	$19.6 \pm 1.0$

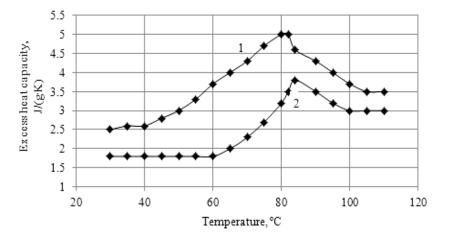


Fig.1. DSC thermogram for agar-agar aqueous solution: 1 – the Panreac agar-agar; 2 – the Helicon agar-agar.

The basic structure of carrageenan is made up of disaccharide units of 3.6-anhydro-D-galactose linked to hemi sulfated D-galactose. The varieties of carrageenan differ in the degree of their disaccharide units' sulphation. The degree of sulphation decreases from iota-carrageenan (two sulphate groups per dissacharide) to kappa-carrageenan (one sulphate group). Higher levels of ester sulfate lower the solubility temperature of the carrageenan producing less strong gels, and thus, contribute to gel inhibition (lambda-carrageenan).

The carrageenans samples concerned varied in the extent of their water-solubility. Kappa-carrageenan dissolved freely in water at 25–30°C, while iota-carrageenan was only slightly-soluble and formed dispersion.

Fig. 2 shows DSC thermograms for kappa- and iota-carrageenan aqueous solutions. Table 11 demonstrates the thermodynamic characteristics ( $T_t$ ,  $\Delta H_t$ ) of the carrageenans under investigation. Kappa-carrageenan displays narrow cooperative endothermic transition within a range of 20–50°C, which agrees with double helix-to-random coil change. Re-scanning of kappa-carrageenan solution (Fig. 1) confirmed the conformational process to be reversible.

For iota-karrageenan we observed a different picture. The first scanning lacked the cooperative calorimetric effect, the polysaccharide being sparingly soluble at room temperature. After heating to  $100^{\circ}$ C we were able to register cooperative endothermic phase transition within  $20-40^{\circ}$ C range. Enthalpy value of iota-carrageenan transition is significantly smaller than the same for kappa-carrageenan (Table 11). This is indicative of a low enthalpy during gel network units' disintegration; in other words, when cooled down the iota-carrageenan solution form a soft, weak gel. The re-scanning (Fig. 2.) shows the reversibility of endothermic phase transition for iota-carrageenan.

Thermodynamic characteristics of carrageenans' aqueous solutions indicate that kappa-carrageenans form more stable gels which have higher melting points. It should be noted that once potassium ion concentration increases in the system, so does temperature of endothermic transition. Thus, the concentration level KCl in 1 M provides for a transition temperature of 80°C and above. This permits the broad-scale adjustment of kappa-carrageenan gel melting points by adding potassium ions into the solution.

That the kappa-carrageenan sample dissolves freely at 25-30°C and forms sufficiently strong gel is indicative of the potassium ion impurities presence that are left behind after extracting kappa-carrageenan from algae.

Actually, iota-carrageenans are water-insoluble at room temperature. High density of negatively charged ions (due to presence of two sulfate groups on iotacarrageenan polysaccharide units) does not permit the successful aggregation of polysaccharide chains. To transform iota-carrageenan solution into gel would involve calcium ions. The ions would shield negatively-charged sulphate groups and cross-link biopolymer molecules ensuring the formation of double-helix.

Summing up the thermodynamic data observed, we can conclude that due to its ability to form thermoreversible gel at room temperature, kappa-carrageenan sample is advisable for creating new functional gel systems based on natural polysaccharides. Moreover, kappa-carrageenan gel strength and temperature can be adjusted by changing potassium chloride concentration. This fact has important implications in developing innovative biodegradable polymers.

Chemically, hydroxypropyl methylcellulose (HPMC) samples differ in the degree of their polymer chain branching, carboxyl group methylation, as well as the methylated group distribution along polymer molecule. HPMC structural features allow its molecules to retain and combine water, form gels, as well as interact with metal cations and proteins. HPMC is used extensively in food industry due to its gel-forming properties.

The HPMC samples studied are freely watersoluble and form stable viscous solutions.

Fig. 3 shows DSC thermogram for hydroxypropyl methylcellulose aqueous dispersions. Thermodynamic characteristics of these dispersions are illustrated in Table 12.

After analyzing Fig. 3 and Table 12 data, we were able to see narrow cooperative endothermic transition from 42 to 44°C in 1.0% solutions for both HPMC samples, which agrees with double helix – to - random coil type transition. After re-scanning, we noted that conformational change for the HPMC solutions is reversible.

Hence, it is advisable to employ 1.0% aqueous dispersions of both hydroxypropyl methylcellulose samples in biodegradable polymers production as compound components.

Ultimately, drawing upon the organoleptic, physical, chemical indicators, as well as chemical and microbiological safety indicators of the Pancreac (Germany) and Helicon (USA) agar-agars, the Bos Science (USA) and Newgreen Pharmachem Co. (China) carrageenans, and the Acros (Belgium) and Ashland Aqualon Functional Ingredients (USA) hydroxypropyl methylcellulose samples, we can conclude the following:

 each of the agar-agar samples conforms with GOST 16280-2002 requirements;

- each of the carrageenan samples conforms with the international food standards CODEX Alimentarius provisions, a CAS 11114-20-8 standard for carrageenans;

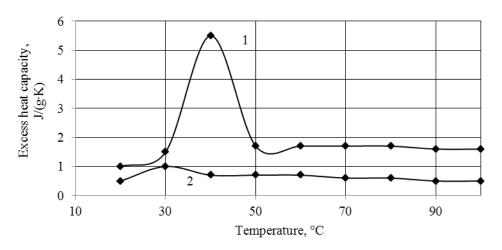
– each of the HPMC samples conforms with the international food standard CODEX Alimentarius provisions, a CAS 9004-65-3 standard for hydroxypropyl methylcellulose.

The thermodynamic analysis of natural polysaccharides aqueous solutions revealed that agaragar and hydroxypropyl methylcellulose samples do form stable gel systems and have much to recommend them in biodegradable polymers production. Among carrageenans of interest is kappa-carrageenan that forms a thermally reversible gel at room temperature.

Taking into account all of the above, the agar-agar, hydroxypropyl methylcellulose and kappa-carrageenan tested appear to be a desirable raw material in future development of biodegradable polymers production technologies.

Table 11. Thermodynamic characteristics of melting the carrageenans' aqueous dispersions

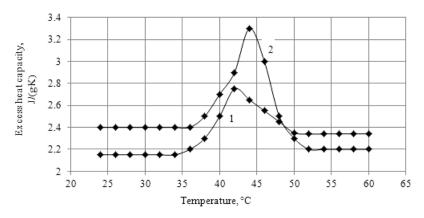
Sample	T <sub>t</sub> , ℃	$\Delta H_t$ , J/g
The Boc Sciences kappa-carrageenan	$42.0 \pm 2.1$	$32.2 \pm 1.6$
The Newgreen Pharmchem Co. iota-carrageenan	$31.2 \pm 1.6$	$5.3 \pm 0.3$



**Fig. 2.** DSC thermogram for carrageenans aqueous dispersions: 1 – Boc Sciences kappa-carrageenan; 2 – the Newgreen Pharmchem Co. iota-carrageenan.

Table 12. Thermodynamic characteristics of melting the aqueous dispersions of hydroxypropyl methylcellulose

Sample	T <sub>t</sub> , ℃	$\Delta H_t$ , J/kg
The Acros hydroxypropyl methylcellulose	$42.5 \pm 2.1$	$7.7 \pm 0.4$
The Ashland Aqualon Functional Ingredients hydroxypropyl methylcellulose	$43.9\pm2.2$	$8.2 \pm 0.4$



**Fig. 3.** DSC thermogram of hydroxypropyl methylcellulose aqueous dispersions: 1 – the Acros HPMC; 2 – the Ashland Aqualon Functional Ingredients HPMC.

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